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Biology of *Maruca vitrata* (Lepidoptera: Crambidae), a serious pest of cowpea and other legume crops: A review

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ABSTRACT

Legume flower and pod borer, *Maruca vitrata fabricius* is a key pest of cowpea and other legume crops, causing up to 80 percent yield loss. The insect is most commonly found in tropical and subtropical areas with wide host range and favorable climate. There are about 39 host plant species of legume crops and weeds and undergoes complete metamorphoses with five destructive larval stages, attacking the flowers, pods, leaves and other parts of the host plant. Many control methods are attributed to its control but most farmers rely largely on chemical pesticides due its physical and immediate action, despite its huge negative effect. However, other alternative such as biocontrol e.g. parasitoid (such as *Apanteles taragamae*), entomopathogenic viruses (e.g. *M. vitrata* Multi-Nucleopolyhedrovirus), and entomopathogenic fungi have recently gained interest and proven effective. Moreover, use of sex pheromones and traps plus cultural control practices such as intercropping, weeding, time and density of planting are also highly efficient and complementary. More data on biology, population ecology, pesticides resistance techniques and natural enemies of *M. vitrata* and their integration would help establishes a tough control method and increased legume crops productivity.

Keywords: Biology, *Maruca vitrata*, pest, cowpea, legume crops

INTRODUCTION

Maruca vitrata Fabricius (Lepidoptera: Crambidae), a legume pod borer has been investigated as destructive pest of cowpea and other legume crops in Africa and Asia [1]. This insect pest is widely distributed throughout tropical and subtropical areas [2], especially where the populace depends largely on agriculture [3]. Previous studies have suggested *M. vitrata* to originate most likely from Indo-Malaysian region [4]. However, present-day studies revealed that the genus *Maruca* might have spread to Africa more recently [5-6] and has already established on 39 species of leguminosae [7]. The larval stage is the damaging stage, attacking flower buds, flowers and young pods [8]. Up to 80 percent cowpea yield losses are recorded [9], especially with little or no control efforts. In West Africa, *M. vitrata* has been identified to form one of a complex of damaging insect pests of cowpea, comprising of *Aphis craccivora* (aphis), *Ootheca mutabilis* (foliage beetles), *Megalurothrips sjostedti* (thrips) and *Clavigralla tomentosicollis* Stål [10]. But the majority of damage is credited to *Maruca vitrata* according to [11].

Considering the above, this paper reviews literature in relation to the biology and other important aspect of *Maruca vitrata* that might help in reducing its effects on legume crops and maximize cowpea production. As this would facilitate hunger alleviation in tropical and subtropical areas [12].

Taxonomy

Maruca vitrata Fabricius is the scientific name of this legume flower and pod borer, synonymously called *Maruca testulalis* Geyer and *croshipora testulalis* Geyer [13]. It belongs to the family Crambidae (order Lepidoptera) [14] formally placed under Pyralidae, which has only a single L seta on its ninth abdominal segment.

Distribution

Several literatures have identified *M. vitrata* to be widely distributed most especially sub-Saharan Africa with wide-ranging host plant (Legumes) and favorable climatic condition [15]. According to recent studies the geographical range of *M. vitrata* has extended to some parts of Europe probably by means of pod transportation [16]. About 39 host species of this pest were identified feeding on flowers, pods, leaves and shoots [17-18]. Despite having such a wide range of alternative hosts in West Africa, *M. vitrata* appears to be migratory along the coastal areas to the dry savanna parts (South-North gradient) [18].

Biology

Eggs are laid singly or in batches of 2-6 [8] on the under surface of leaves, terminal shoots and flower buds [19]. The freshly laid eggs are milky white in color, oval in shape, dorsoventrally flattened and stuck to its substrate [19]. Temperature effect the number and period of egg laying [20] and up to 400 eggs are laid in batches of 2-16 [21] while the incubation period varies from 2.54 ± 0.04 days [19].

There are five larval instars lasting between 8-10 days [19] or even up to 16 days [15] depending on climatic condition and the host plants. A shorter duration was observed [8] who reared the larvae with artificial diet. Larval feed component also affects the biology of the insect and it preferentially feed on the reproductive organs of the host plant [8] for about three week and then migrates to the pods before pupation. Larval body is semitransparent and spotted on each segment and the spotting intensity varies [22] and the spots fade before pupation [23].

The pupae are elongated, measuring about 13 mm in length and with shouldered appearance [24]. Early pupal stage is greenish but turns brown when fully developed [22] and concealed in a cocoon on dry leaves, flowers and other dead plant matters [23]. The pupal period is normally one or two weeks [22]. No sign of diapause during the dry season [25].

The Adult are medium sized [22] and both sexes are morphologically alike. The forewings are brown having white spot and black-edged while the hind wings are semi-hyaline [22]. The highest percentage of mating and oviposition is in 4 or 5 nights of pairing [21] and the suitable temperature range for this is between 20 °C and 25 °C with 80 % and above humidity [26]. Mating occurs around 20:00 h and 05:00 h and the males and female longevity of 7-10 days and 5-6 days respectively [22].

Economic Importance

Several findings have described *M. vitrata* as the most devastating insect pest causing poor yield and considerable losses of cowpea worldwide [1]. The larval stage is the damaging stage [8] and it does so by entering the buds and seed pods. The damaged pods are completely or partially eaten out and entrance also allows water into the pod and stains the left behind seeds. The buds, flowers and leaves are also damage [27] which may be consumed and or bound together by the larvae. The young larvae usually feed on and damage the flower while the older ones feed most often on the pod [28].

Control**Chemical Control**

To manage *M. vitrata* and other insect pests, farmers resort to many methods based on their knowledge and financial status [29]. Despite the financial burden and other problems associated with chemical insecticides, copious data have revealed that they are the most widely used [30]. Recent studies conducted in Thailand revealed that more than 90% of the surveyed growers relied on chemical pesticides and two-thirds of them applied it once a week [31]. Likewise, application of insecticidal chemicals once at both flowering and podding stages greatly increase grain yield [32]. Pesticides such as methomyl, cypermethrin, endosulfan, dimethoate, carbaryl, lambda-cyhalothrin 5 EC, beta-cyfluthrin and monocrotophos have all proven effective against *M. vitrata* [33-35] if sprays regularly. Dimethoate combined with cypermethrin gives efficient control in cowpea field [36]. A combination of deltamethrin and

lambda-cyhalothrin mix with dimethoate has also shown activity against this insect pest [32] especially when applied prior to pods infestation.

Chemical pesticides in the short-term exhibit significant control, increased yields, immediate effect and plant health improvement that easily attract less-literate farmers [37], but in the long run, causes serious long-term effects such as insecticide resistance and low level of tolerance in *M. vitrata*, which has already been identified [33]. For instance, resistance to cypermethrin, endosulfan and dimethoate were detected and non-chemical control or its use only when necessary was recommended [33]. Other effects include; wiping out of natural enemies [38], pollution of the environment, toxicity to living organisms, farmers, consumers and are costly as well [39].

Biological Control

In recent years, biological control practices, an alternative method to chemical pesticides [40], has gained interest in the fight against insect pest [5] which has to some extent, been proved to be a better control method. This practice includes; the use of natural enemies (parasitoid e.g. *Phanerotoma leucobasis*, *Pristomerus* sp., *Testudobracon* sp. *Apanteles taragamae*) of eggs and larvae of *M. vitrata* [18, 6], entomopathogenic viruses e.g. *M. vitrata* Multi-Nucleopolyhedrovirus [41-42] and entomopathogenic fungi. *M. anisopliae* and *B. bassiana* Isolates [43]. *Bacillus thuringiensis* subsp. *aizawai* has also been observed to be highly effective [44]. Earlier studies have also revealed *M. vitrata* susceptibility to *Bacillus thuringiensis* δ -endotoxins in tropical and subtropical areas [45]. Most of these methods have shown encouraging results but some have not yet been accepted and more studies at population-level are vital for fruitful biocontrol implementation [46].

Botanical control

Many potentialities of botanicals against *M. vitrata* have recently been documented. About 90 percent larval mortality to neem concentration of 50,000 ppm has been reported [44]. Similarly, a laboratory experiment by [47] revealed *Allium sativum* bulb, *Piper guineense* and *Azadirachta indica* seed extracts as effective in egg hatch reduction with black pepper and garlic bulb having the highest reduction at all concentrations. In another study, [48] examined Neem, papaya and Hyptis species to be effective on both *Maruca* and other pest like *Thrips* with neem having better result. Other findings on neem oil also revealed its effectivity against *M. vitrata* in India and Africa [49]. Equally, [50] found that, neem seed kernel extract (5 percent) with Dichlorvos (0.5 ml per liter of water) produced an excellent result.

Cultural Control

Cultural practices such as intercropping, weeding, time and density of planting lessen the damage in cowpea [28]. A plant spacing range of 1.0–1.5m can reduce *M. vitrata* infestation and most other yield-limiting insect infestation [51] and planting at 30x20 cm² or 60x20 cm², at the onset of the rain also gives a better result [52]. Furthermore, [53] reported an increased bird perching resulting from intercropping with sorghum seeds and summer ploughing reduces pod borer by 85 % and increase legume crops productivity. Daily inspections and hand picking of the eggs and larvae of *M. vitrata* are by far better than the use of synthetic chemicals when the plants are few.

Sex Pheromones and Traps

The use of sex pheromones and traps have established a high degree of efficiency and proved complementary in the control of *M. vitrata* population in recent times. Most studies on sex pheromone of insect pests of legumes were conducted on *M. vitrata* [54]. Existing research have identified (E,E)-10,12-hexadecadienol, and (E)-10-hexadecenal as minor components [54] and (E,E)-10,12-hexadecadienal as major component [55] of sex pheromones which are both effective. Additionally, two more components, (E)-10-hexadecenol and (Z,Z,Z,Z,Z)-3,6,9,12,15-tricosapentaene were identified with increased catch [56].

Lots of these moths (up to 1500 moths in just a night) were caught during cowpea growing season in Kano, Nigeria, with a light trap [57] which indicates that it can complement other methods to reduce *M. vitrata* damage to cowpea field.

CONCLUSION

Going by its recent distribution and development of resistance to some chemical pesticides, *M. vitrata* is becoming a potential threat to dwindling global cowpea and other legume production. Thus, *M. vitrata* requires a great deal of attention. More comprehensive, diverse and up-to-date information on its ecology, diversity, patterns of migration,

pesticides resistance techniques and off-season occurrence are needed to understand it better and screen for the most appropriate management strategies. Identification of more natural enemies of this pest and its integration with cultural practices and other biocontrol strategies would be an effective and safer control approach. Sex pheromones and traps are also effective in reducing the population pressure this pest. Intensive research into the components of the sex pheromone of this insect pest in different geographical regions is still required to fine-tune the pheromone based control of *M. vitrata* populations. Crop improvement programs should focus on producing resistant and genetically transformed yet easily accessible cowpea and other legumes seeds to local farmers.

REFERENCES

- [1] Baoua I., Ba N. M., Agunbiade T. A., Margam V., Binso-Dabire C. L., Sanon A. & Pittendrigh B. R. *International Journal of Tropical Insect Science*, **2011**, 31, 212–218.
- [2] Liao, C. T. & Lin, C. S., *Plant Protection, Bulletin*. **2000**, 42: 213-222.
- [3] World Bank, *World Development Report 2008: Agriculture and Development*. Washington, DC. World Bank, 2007, <http://openknowledge.worldbank.org/handle/10986/5990> Licence: CC by 3.0 IGO.
- [4] CAB International, *Crop Protection Compendium*, **2005** Edition. Wallingford, UK: CAB International, 2005, accessible at; www.cabicompendium.org/cpc.
- [5] Tamò M., S. Ekesi, N. K. Maniania, & A. Cherry, *CABI Publishing*, Wallingford, **2003**.
- [6] Srinivasan, R., M. Tamò, PA-C. Ooi, & W. Easdown, *Biocontrol News and Information*, **2007**, 28: 34–37.
- [7] Arodokoun, D. Y., Tamò, M., Cloutier, C. & Adeoti, R., *Insect Science Application*, **2003**, 23, 103–113.
- [8] Traore, F., C. L. Dabire-Binso, N. M. Ba, A. Sanon & B. R. Pittendrigh, *Memoirs of the Faculty of Agriculture, Kagoshima University*, **2013**, 38: 41-44.
- [9] Dannon E. A., PhD thesis, Wageningen University, Netherlands, **2011**, 188 Pages.
- [10] Dabire-Binso, C. L., Ba, N. M., Sanon, A., Drabo, I. & Foua Bi K., *International Journal of Tropical Insect Science*, **2011**, 30, 192-199.
- [11] Egho, E. O. & Emosairue, S. O. *Agriculture and Biology Journal of North America*, **2010**, 1, 5, 938-945
- [12] IITA, *Improved cowpea varieties hit Nigeria's savannas*, **2010**, [Online] Available from: http://old.iits.org/cm/details/news_details.aspx?articleid=3488&zoneid=81 [Accessed 1st May 2012].
- [13] Ayodele, M. & Kumar, L., *Crop gene bank knowledge base: insect-cowpea*, 2011, (Online) available from: http://croptgenebank.sgrp.cgiar.org/index.php?option=com_content&view=article&id=57&Itemid=729&lang=es
- [14] Solis, M. A. *Proceedings of the Entomological Society of Washington*, **1999**, 101 (3):645-686.
- [15] Shanower, TG., Romeis, J. & Minja, EM., **1999**. *Annual Review of Entomology*, 44: 77-96
- [16] UKmoths, 1401 *Mung moth*, 2013, [Online] available from: <http://ukmoths.org.uk/show.php?bf=1401>
- [17] Sharma, H. C., Saxena, K. B. & Bhagwat, V. R., *Information bulletin* 55. Patancheru, India, International Crops Research Institute for the Semi-arid Tropics (ICRISAT), **1999**.
- [18] Arodokoun, D. Y., Tamò, M., Cloutier, C. & Brodeur, J., *Agriculture, Ecosystems & Environment*, **2006**, 113: 320–325.
- [19] Naveen V., Naik, M. I., Manjunatha M., Shivanna B. K. & Sridhar S., *Karnataka Journal Agricultural Science*, **2009**, 22, 668-669.
- [20] Chi, Y., Sakamaki, Y. Tsuda, K. & Kusigemati, K., *Japanese Journal of Applied Entomology and Zoology*, **2005**, 49, 29–32.
- [21] Jackai, L. E. N., Ochieng, R. S. & Raulston, J. R., *Entomologia Experimentalis et Applicata*, 1990, 59, 179-186.
- [22] Ganapathy, N. *Madras Agriculral Journal*, **2010**, 97, 7-9, 199-211
- [23] Arulmozhi. K. M.Sc. (Ag.) *Thesis*. Tamil Nadu Agricultural University, Coimbatore, **1990**, 117 Pages.
- [24] Passoa, S. And D. Bean. Eastern Pest Survey Committee Guidelines for the Cooperative Agricultural Pest Survey Program, **2001**, 5 pages.
- [25] Arodokoun, D. Y., M. Tamò, C. Cloutier & R. Adeoti, *Insect Science and its Application* 2001 (submitted).
- [26] Abdullahi M. M, *PhD Thesis*, university of Greenwich, England, **2013**.
- [27] Huang C. C, Peng W. K, Talekar N. S., *Biological Control* **2003**, 48, 407–416.
- [28] Sharma, H. C., *Crop Protection*, **1998**, 17: 373–386.
- [29] Rosendahl, I. V., Laabs, C., Atcha-Ahowé, B., James & W. Amelung, *Journal of Environmental Monitoring*, **2009**, 11, 6, 1157–1164.
- [30] Alam, S. N., Ahmed, G. J. U., Khorsheduzzaman, A. K. M., Karim, A. N. M. R. & Rajotte, E. G., 5th National IPM Symposium, April 4–6, **2006**, accessible at

<<http://www.ipmcenters.org/IPMSymposiumV/posters/index.html>>.

- [31] Bhattarai, M., Patricio, R., Yule, S., Wu, M. H. & Srinivasan, R., 7th ASAE Conference “Meeting the Challenges Facing Asian Agriculture and Agricultural Economics Toward a Sustainable Future,” October 13–15, **2011**, Hanoi, Vietnam, 2011.
- [32] Kamara, A. Y., Chikoye, D., Ornoigui, L. O. And Dugje, I. Y., *Journal of Food Agriculture and Environment*, **2007**, *5*, 154-158.
- [33] Ekesi, S., *International Journal of Pest Management*, **1999**, *45*, 57–59
- [34] Dushyant K. & Biswajit Das Pal, D. P., *Environment and Ecology*, **2006**, *24*, 1, 184-186.
- [35] Mohapatra, S. D. & Srivastava, C. P., *Indian Journal of Entomology*, **2008**, *70*, 1, 61-63.
- [36] Dzemo, W. D., Niba, A. S. and Asiwe, J. A. N., *African Journal of Biotechnology*, **2010**, *9*, 11, 1673-1679
- [37] Singh S. R, L. E. N Jackai, J. H. R dos Santos and C. B. Adalla, John Wiley and Sons Ltd, Nigeria, **1990**.
- [38] Heong, K. L., G. B. Aquino and A. T. Barrion, *Crop protection*, **1992**, *11*, 371–379.
- [39] Sharah, H. A & Ali, E. A., *International Journal of Agriculture and Biology*. **2008**, *10*, 255 – 260
- [40] Nicholson, G. M., *Toxicon*, **2007**, *49*, 413-422.
- [41] Lee, S-T., Srinivasan, R., Lo, Y-J. & Talekar, N. S., *BioControl* **2007**, *52*, 801–819.
- [42] Tamò, M, I. Godonou, J. Braima, R. Srinivasan, D. Kpindu, C. Kooyman, C. Agboton, B. Datinon, S. Nakamura, T. Adati, N. Maniania, & S. Ekesi. *International Plant Protection Congress*, Honolulu, 6–11 August **2011**.
- [43] Ekesi, S., Adamu, R. S. & Maniania, N. K., *Crop Protection*, **2011**, *21*, 589–595.
- [44] Yule, S. and R. Srinivasan, *Journal of Asia-Pacific Entomology*, **2013**, *16*, 357–360
- [45] Srinivasan R., *Journal of Invertebrate Pathology*, **2008**, *97*, 79-81; PMID: 17689558; <http://dx.doi.org/10.1016/j.jip.2007.06.005>.
- [46] Margam, VM., Coates, BS., Hellmich, RL., Agunbiade, T., Seufferheld, MJ., Sun, W., Ba, MN., Sanon, A., Binso-Dabire, CL., Baoua, I., Ishiyaku, MF., Covas, FG., Srinivasan, R., Armstrong, J., Murdock, LL. & Pittendrigh, BR., *PLOS ONE*, **2011**, *6*, 2, e16444.
- [47] Ekesi, S., *Phytoparasitica*, **2000**, *28*, 305-310.
- [48] FAO/TECA Pheromone traps for the management of the cowpea pest *Maruca Vitrata*. Published in *TECA* 2012, (<http://teca.fao.org>)
- [49] Jackai, L. E. N., & Oyediran, I. O., *Insect Science Application*, **1991**, *12*, 103–109.
- [50] Gopali, J. B., Raju T., Mannur, D. M. and Suhas Y., *Karnataka Journal of Agricultural Science*, **2010**, *23*, 35-38.
- [51] Asiwe, J. A. N., S. Nokoe, L. E. N. Jackai, & F. K. Ewete, *Crop Protection*, **2005**, *24*, 465–471
- [52] Karungi, j., Adipala, E., Kyamanywa, S., Ogenga-Latigo, M. W., Oyobo, N. and Jackai, L. E. N., *Crop Protection*, **2000**, *19*, 4, 237-245
- [53] Sandeep, K., S. K. Kannaujia, A. V. and V. K. Singh, *Plant Archives*, **2013**, *13*, 1, 171-172.
- [54] Downham M. C. A, Hall D. R., Chamberlain D. J, Cork A., Farman D. I., Tamo M., Dahounto D., Datinon B., Adetonah S., *Journal of Chemical Ecology*, **2003**, *29*, 989–1012.
- [55] Adati, T., Tatsuki, S., *Journal of Chemical Ecology*, **1999**, *25*, 105–116
- [56] Hassan M. N., PhD Dissertation submitted to University of Greenwich, UK, **2007**, 244 Pages.
- [57] Bottenberg, H., Tamò, M., Arodokoun, D., Jackai, L. E. N., Singh, B. B & Youm, O., *Advances in Cowpea Research*. Ibadan, Nigeria, International Institute of Tropical Agriculture (IITA) and Japan International Center for Agricultural Sciences (JIRCAS), IITA, **1997**, 271–284.